

## COMPARISON OF CHEMICAL COMPOSITION OF FRESH AND FERMENTED CABBAGE JUICE

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### Abstract

White cabbage (*Brassica oleracea* var. *capitata*) is an affordable and available vegetable in local markets around the globe. It is a source of vitamins, micro and macro nutrients. The aim of this study was to compare the chemical composition of fresh and fermented cabbage juices from three different varieties. In this study vitamin C content, antiradical activity, total phenolic content, and total carotenes were determined. Results show a significant ( $p < 0.05$ ) influence of cabbage variety on vitamin C content. Fermentation process decreased vitamin C content in the variety 'Selma' but increased in 'Ramkila' and 'Kilpatons'. There were no significant differences between varieties ( $p > 0.05$ ) in the antiradical activity (by DPPH method) of fresh cabbage juice while fermentation process slightly increased it in varieties 'Ramkila' and 'Kilpatons' but significantly increased it in 'Selma' (from 96.66 for fresh to 189.54 mg 100 g<sup>-1</sup> on dry weight (DW) for fermented). There was no consistency in the antiradical activity by ABTS<sup>+</sup> method. Fermentation process slightly decreased it in variety 'Ramkila', significantly decreased it in variety 'Selma' (805.72 for fresh to 356.76 mg 100 g<sup>-1</sup> DW for fermented) but significantly increased in 'Kilpatons' (517.09 for fresh to 845.48 mg 100 g<sup>-1</sup> DW for fermented). Fermentation process significantly influenced contents of total phenolic compounds in two varieties – 'Ramkila' (1176.1 for fresh to 1637.7 mg 100g<sup>-1</sup> DW for fermented) and 'Kilpatons' (1106.3 for fresh to 1872.9 mg 100g<sup>-1</sup> DW for fermented). Results showed that white cabbage or sauerkraut is not a beneficial source of carotenes.

**Keywords:** fermented cabbage, antiradical activity, total phenolic content

### Introduction

White cabbage (*Brassica oleracea* var. *capitata*) is an affordable and available vegetable in local markets around the globe. Annually cabbage and brassica vegetables are consumed approximately 6.3 kg worldwide (Rokayya et al., 2013) and more than 8 kg of white cabbage in Latvia per capita (Gailīte, 2018).

It is a vegetable of low caloric value (24–36 kcal 100 g<sup>-1</sup>), low protein (1.4 g 100 g<sup>-1</sup>), low fat (0.2 g 100 g<sup>-1</sup>) content, but high in minerals (such as potassium: 208 mg 100 g<sup>-1</sup>, calcium: 46 mg 100 g<sup>-1</sup>, magnesium: 12 mg 100 g<sup>-1</sup>, vitamin C (329.5 mg 100 g<sup>-1</sup>), vitamin A (31 µg 100 g<sup>-1</sup>), dietary fibre (3.0 g 100 g<sup>-1</sup>) and water content (92 g 100 g<sup>-1</sup>). Cabbage contains moderate carbohydrate (glucose: 2.0 g 100 g<sup>-1</sup>, fructose: 1.8 g 100 g<sup>-1</sup>) amounts, and have a high level of phenolic compounds including polyphenols (Rodriguez-Amaya, 2015). β-carotene content in cabbage ranged from 0.009–0.124 mg 100 g<sup>-1</sup> fresh weight, total phenolics: 12.58–34.41 mg 100 g<sup>-1</sup> fresh weight (Singh et al., 2006). Total phenolic contents, antioxidant capacity, flavonoid content are influenced by many factors, including growing developmental stages (Samec, 2011). Most common way to use cabbage is either fresh in soups and salads, or fermented – sauerkraut.

Sauerkraut fermentation is a dynamic biochemical system, where chemical composition and microbial ecology of the system are continuously changing (Lu et al., 2003). To ferment cabbage, varieties with the highest carbohydrate content with a sugar content of at least 4% are selected. Fermentation process using lactic acid bacteria in cabbage increases the content of vitamins, free amino acids and other physiologically active substances in the product, though the choice of starter and cabbage variety has a noticeable impact on chemical composition (Vatansever, 2017; Martinez-

Villaluenga et al., 2012). Spontaneous sauerkraut fermentation relies on a small population of lactic acid bacteria (LAB) (which are naturally present on fresh vegetables) and their metabolites. The process is divided into heterofermentative and homofermentative stages. In the first stage, the activity of heterofermentative *Leuconostoc mesenteroides* determines the quality of sauerkraut. Acid-tolerant *Lactobacillus* species takes over second homofermentative stage with decrease in pH (4.5–3.5) and accumulation of lactic acid. (Yoon et al., 2002; Lu et al., 2003). Consumption of sugars, pH reduction and acid production is very rapid in the first stage (~ first 2 weeks) and slows thereafter and remains unchanged after day 30 (Yoon et al., 2002). Strains isolated during the first stage (day 1 to day 3) belong to genus *Leuconostoc* or *Weisella*, after that it is *Lactobacillus* strains (*L. plantarum* and *L. brevis*) (Yoon et al., 2002).

Biochemical tests showed that *Leuconostoc* and *Weisella* strains produce carbon dioxide from glucose as well as characteristic slime of dextran from sucrose. None of *Lactobacillus* (*L. plantarum* nor *L. brevis*) strains produce slime colonies on sucrose agar, and only *L. brevis* strain produces carbon dioxide (Lu et al., 2003). To ensure the correct sequence of LAB species, which is essential to achieve a stable product with typical flavour and aroma, producers choose to use LAB starter culture (*Leuconostoc mesenteroides*) as well as to reduce NaCl content as low as 0.6% (Viander et al., 2003).

Bacterial growth is involved in the metabolism of phenolic compounds and is compound dependent (Rodriguez et al., 2009; Alberto et al., 2012).

Fermentation process leads to formation of bioactive compounds (Palani et al., 2016). The influence of fermentation process on chemical composition in white cabbage has been done by Spanish researchers. It states

that fermentation process increases ascorbic acid content, antioxidant activity and nitric oxide production inhibitory activity, though the choice of starter has a noticeable impact (Martinez-Villaluenga et al., 2012). For ascorbic acid formation, initial glucobrassicin levels in vegetables are important and is a reason why the amount of ascorbigen can significantly vary between different varieties (Wagner, Rimbach, 2009). The titratable acidity in sauerkraut ranges between 0.9–1.5% (Trail et al., 1996).

Consumption of fresh and fermented cabbage in Latvia is considerable. However little or no research was done to compare cabbage varieties grown in Latvia and how fermentation process affects their chemical composition.

The aim of this study was to compare chemical composition of fresh and fermented cabbage juices from three different varieties.

### Materials and Methods

Experimental work was carried out in Latvia University of Life Sciences and Technologies, Faculty of Food technology.

#### Sample preparation

Three varieties of fresh cabbage heads were delivered by a farmer. Three 1 L bottles of sauerkraut juice from the same varieties were delivered by the same farmer. The varieties were: 'Selma' (with a light green firm head), 'Kilpatons' (with a greener, not so firm, head; sweet to taste), and 'Ramkila' (a big, very light, and very firm head with some spoiled leaves in between).

Cabbage heads were cleaned from outer, not-fresh leaves. The spoiled inner leaves of variety 'Ramkila' were removed. Segments (3 cm wide) were cut out of each head and juice was extracted with masticating slow juicer (Easyline, Villa-Verucchio model ELCJE6203235M).

#### Chemical and physical analyses

*Vitamin C* was determined using iodine method T-138-15-01:2002 (Segliņa, 2007) which determines reduced form of ascorbic acid (L-ascorbic acid). Experimental samples were mixed with 100 mL 6% H<sub>2</sub>C<sub>2</sub>O<sub>4</sub> solution blended and filtered. 10 mL of filtrate was mixed with 2 mL of 1% starch solution and titrated with 0.05 M iodine solution.

*Preparation of extracts.* Samples of fresh and fermented juices (~ 10 g) of each variety were extracted with 20 mL of 80% ethanol by stirring on the magnetic stirrer for 45 minutes. Two repetitions were made. Ethanolic extracts were then filtered into 25 mL flasks and stored at 4±2 °C till further analysis. Extracts were used for the estimation of total phenolic contents and antiradical activities (DPPH; ABTS<sup>+</sup>) (Rokayya et al., 2013).

*Total phenolic content* in extracts was estimated spectrophotometrically using Follin-Ciocalteu reagent according to Prasad et al. (2013) with some modifications. Three repetitions of 0.5 mL of extract were mixed each with 2.5 mL Follin-Ciocalteu reagent (diluted ten times with deionized water), left to react for

5 min. Then 2 mL 7.5% Na<sub>2</sub>CO<sub>3</sub> were added and mixtures were left to react for another 30 min. The absorption was read spectrophotometrically at 765 nm on Jenway 6300 (Baroworld Scientific Ltd., UK). The total phenolic content was determined using standard gallic acid calibration curve and results were expressed as milligrams of gallic acid equivalent (mg GAE 100 g<sup>-1</sup> DW).

*Antiradical activity* was determined using ABTS<sup>+</sup> (2,2-azino-di-3-ethylbenzothiazoline-sulphonic acid) decolouration method and DPPH (2,2-diphenyl-1-picrylhydrazyl) radical scavenging assay. The ABTS<sup>+</sup> stock solution was made with addition of potassium persulfate as an oxidation agent (Rokayya et al., 2013) and left to react in the darkness for 12 h. To obtain the working solution of ABTS<sup>+</sup>, the stock solution was diluted with phosphate buffered saline (PBS) to the absorption of 0.800±0.030 at 734 nm (against the blank) on JENWAY 6300. Three repetitions of 0.05 mL sample extract were mixed with 5 mL of ABTS<sup>+</sup> solution, left to react for 10 min and absorptions were determined as had been described previously.

The DPPH radical scavenging assay was done according to the method of Kriengsak et al. (2006) with some modifications. The stock solution was made by mixing 0.004 g DPPH with 96% ethanol to the absorption of 1.000±0.02 units against the blank on spectrophotometer at 517 nm. Three repetitions of 0.5 mL sample extracts were mixed with 3.5 mL freshly made DPPH stock solution, left to react in the dark for 30 min and then absorption had been measured.

*Total carotenes* were determined using method described by Kampuse et al. (2015) with modifications. Samples of 5 g (with precision of 0.0002 g) of fresh cabbage and sauerkraut juices were mixed with 20 mL 96% ethanol and stirred magnetically, after 15 min 25 mL of petrol ether (80–110) was added and stirring continued for another 60 min. Samples were left to settle and results were read using UV/VIS spectrophotometer Jenway 6705.

#### Statistical analysis

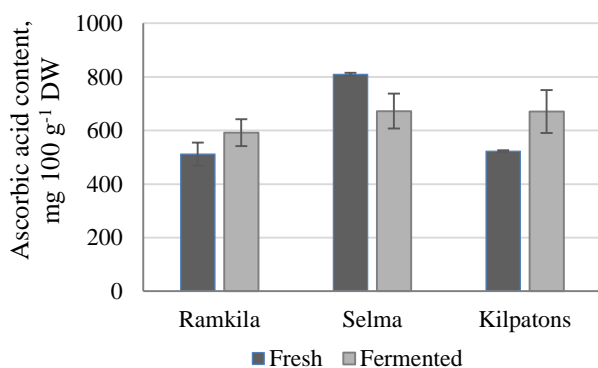
The differences between results were analysed using two-factor analysis of variance (ANOVA) followed by Tukey-Kramer method. The obtained results were presented as means with standard errors. Differences among results were considered to be significant if p<0.05.

### Results and Discussion

Chemical composition of fresh and fermented cabbage juices varied among cabbage varieties which might be due to different initial physical characteristics (Thakur et al., 2017) as well as it is influenced by growing conditions, storing, fermentation process and many more factors (Kusznierewicz et al., 2008; Wagner, Rimbach, 2009; Palani et al., 2016).

*Vitamin C.* Analysing obtained results, it was determined that there is a significant (p<0.05) influence of cabbage variety on vitamin C (Fig. 1) content, also fermentation process affected it differently in studied

varieties. In two of analysed cabbage varieties vitamin C content increased after fermentation process – ‘Ramkila’ (511.8 for fresh to 591.9 mg 100 g<sup>-1</sup> DW for fermented) and ‘Kilpatons’ (522.6 for fresh to 671 mg 100 g<sup>-1</sup> DW for fermented), while variety ‘Selma’ had a decrease in vitamin C after fermentation (809.7 for fresh to 672.4 mg 100 g<sup>-1</sup> DW for fermented). Differences in results can be explained by enzymatic reactions of chemical compounds in varieties as described by Wagner et al. (2009) and Martinez-Villaluenga et al. (2009). According to Thakur et al. (2017), ascorbic acid content increases during fermentation up to day 21, after which it gradually decreases in all varieties.



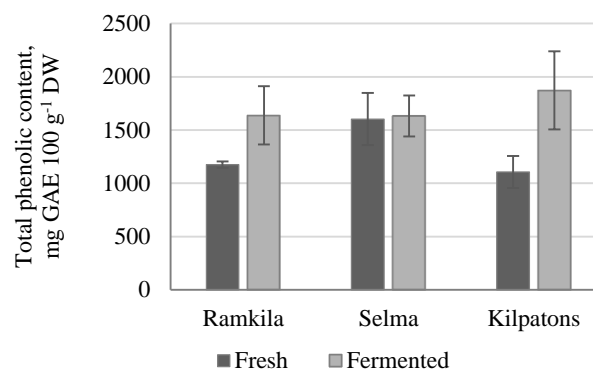
**Figure 1. Ascorbic acid content in fresh and fermented cabbage juice, mg 100 g<sup>-1</sup> DW**

The loss of vitamin C due to cabbage fermentation partially may be explained by ascorbic acid involvement in ascorbigen formation. It also may be influenced by the production process of sauerkraut – vitamin C depletion occur when vegetables are severely cut or shredded. Trimming of outer leaves, that contain more vitamin C than inner leaves, results in greater decrease in vitamin C than enzymatic breakdown by ascorbate oxidase, autoxidation and so on (Martinez-Villaluenga et al., 2009). To avoid decrease of vitamin C in fermentation process, it is advisable to choose fresh cabbage heads with little or no damaged outer leaves and to control the fermentation process, time and temperature.

#### Total phenolic contents

Fermentation process significantly influenced total phenolic contents (Fig. 2) in two varieties – ‘Ramkila’ (1176.1 for fresh to 1637.7 mg 100 g<sup>-1</sup> DW for fermented) and ‘Kilpatons’ (1106.3 for fresh to 1872.9 mg 100 g<sup>-1</sup> DW for fermented). However, total phenolic content in the variety ‘Selma’ was not significantly ( $p < 0.05$ ) influenced by fermentation process (1603.5 for fresh to 1632.4 mg 100 g<sup>-1</sup> DW for fermented). One of the reasons, as Kusznierevicz et al. (2007) have come to conclusion that antiradical activity initially increases during wounding or shredding for spontaneously fermented sauerkraut. Fermentation and chemical processes incurred by lactic acid can induce formation of novel compounds that can neutralize free

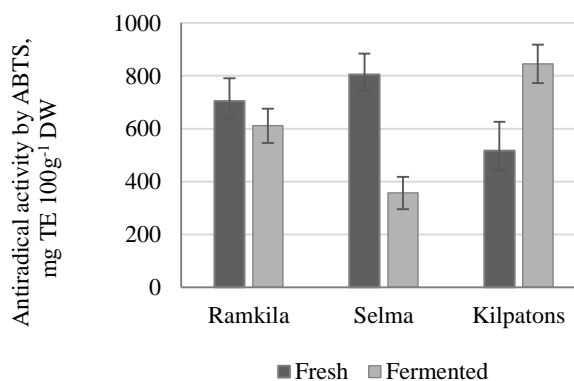
radicals. However, metabolic pathways of biosynthesis or degradation of phenolic compounds by lactic acid bacteria have not been completely described (Rodriguez, 2009).



**Figure 2. Total phenolic content in fresh and fermented cabbage juice, mg GAE 100 g<sup>-1</sup> DW**

#### Antiradical activity

Antiradical activity by ABTS<sup>+</sup> decolouration method showed significant ( $p > 0.05$ ) variations (Fig. 3). Variety ‘Ramkila’ (704.7 for fresh to 611.1 mg 100 g<sup>-1</sup> DW for fermented) and ‘Selma’ (805.7 for fresh to 356.8 mg 100 g<sup>-1</sup> DW for fermented) showed decreasing results, whereas in variety ‘Kilpatons’ antiradical activity increased (517.1 for fresh to 845.5 mg 100 g<sup>-1</sup> DW for fermented).

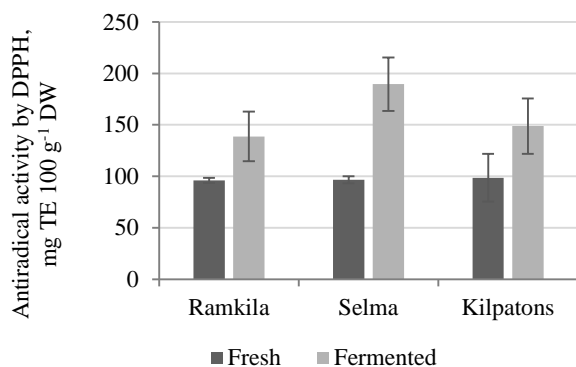


**Figure 3. Antiradical activity by ABTS<sup>+</sup> in fresh and fermented cabbage juice, mg TE 100 g<sup>-1</sup> DW**

There were no significant differences between varieties ( $p > 0.05$ ) in DPPH radical scavenging assay in fresh cabbage juice, (95.9–98.5 mg 100 g<sup>-1</sup> DW). Fermentation process significantly increased the antiradical activity (138.7–189.5 mg 100 g<sup>-1</sup> DW) and there were significant differences between varieties (Fig. 4), too ( $p < 0.05$ ). DPPH method is widely used to determine antiradical / antioxidant activity of purified phenolic compounds (Shalaby, Shanab, 2012). The overall content of phenols tends to increase which also can explain increase of DPPH antiradical activity. As concluded by Kusznierevicz et al. (2007), it is not

always the case that fermenting rises antiradical activity and vitamin C content.

Different antiradical activity results can be explained by differences in radical assay methods. DPPH is sensitive to acidic pH, samples react very slowly not reaching steady state after 8 hours, whereas ABTS<sup>+</sup> method has the extra flexibility to be used at different pH levels, samples react rapidly, reaching steady state within 30 minutes.



**Fig. 4. Antiradical activity by DPPH in fresh and fermented cabbage juice**

ABTS<sup>+</sup> assay measures the relative ability of antioxidant to scavenge the ABTS<sup>+</sup> generated in aqueous phase as compared with a water-soluble vitamin E analogue standard (Trolox) (Shalaby, Shanab, 2012) so it's radicals react with different compounds and show higher antioxidant capacity in cabbage juice (Šamec, 2011). The rise of ABTS<sup>+</sup> antiradical activity in variety 'Kilpatons' could be explained by different and more water-soluble compounds.

#### Total carotenes

Results showed that white cabbage and sauerkraut are not a beneficial source of carotenes, which is in agreement with the information reported by Singh (2006). Variety 'Ramkila' showed a decrease in total carotenes from 9.45 (for fresh) to 3.77 mg 100 g<sup>-1</sup> DW (for fermented) and 'Selma' from 8.28 (for fresh) to 0.27 mg 100 g<sup>-1</sup> DW (for fermented), while variety 'Kilpatons' had an increase from 4.36 (for fresh) to 10.35 mg 100 g<sup>-1</sup> DW (for fermented).

#### Conclusions

Fermentation process positively influenced DPPH radical scavenging activity in tested cabbage varieties, however there is no such trend in the ABTS<sup>+</sup> activity. Fermentation process positively influenced variety 'Kilpatons' – total phenolic contents, vitamin C and antiradical activity (ABTS<sup>+</sup>; DPPH) increased significantly. In variety 'Selma', fermentation process had little influence on total phenolic contents, but antiradical activity by ABTS<sup>+</sup> method and vitamin C content decreased. In variety 'Ramkila' – antiradical activity by ABTS<sup>+</sup> method decreased but total phenolic content and vitamin C increased.

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