

# NIH Public Access

Author Manuscript

Stat Anal Data Min. Author manuscript; available in PMC 2014 February 03.

## Published in final edited form as:

Stat Anal Data Min. 2013 December 1; 6(6): 496–505. doi:10.1002/sam.11196.

## A Weighted Random Forests Approach to Improve Predictive Performance

## Stacey J Winham<sup>1,§</sup>, Robert R Freimuth<sup>1</sup>, and Joanna M Biernacka<sup>1,2,§</sup>

<sup>1</sup>Department of Health Sciences Research, Mayo Clinic, 200 First Street Southwest, Rochester, MN 55905 USA

<sup>2</sup>Department of Psychiatry and Psychology, Mayo Clinic, 200 First Street Southwest, Rochester, MN 55905 USA

## Abstract

Identifying genetic variants associated with complex disease in high-dimensional data is a challenging problem, and complicated etiologies such as gene-gene interactions are often ignored in analyses. The data-mining method Random Forests (RF) can handle high-dimensions; however, in high-dimensional data, RF is not an effective filter for identifying risk factors associated with the disease trait via complex genetic models such as gene-gene interactions without strong marginal components. Here we propose an extension called Weighted Random Forests (wRF), which incorporates tree-level weights to emphasize more accurate trees in prediction and calculation of variable importance. We demonstrate through simulation and application to data from a genetic study of addiction that wRF can outperform RF in high-dimensional data, although the improvements are modest and limited to situations with effect sizes that are larger than what is realistic in genetics of complex disease. Thus, the current implementation of wRF is unlikely to improve detection of relevant predictors in high-dimensional genetic data, but may be applicable in other situations where larger effect sizes are anticipated.

## Keywords

Random Forests; genome wide association; high-dimensional data; genetic data; gene-gene interactions; weighting

## Introduction

In traditional genome-wide association studies, data collected at hundreds of thousands to millions of genetic variants are analyzed univariately in order to determine association with a disease. These tests of association do not account for the effects of other genetic variants, ignore complex models such as gene-gene interactions, and have low power to detect most genetic effects. Thus, identified variants account for a very small proportion of variation in complex traits [1, 2].

To circumvent these issues, data-mining and machine learning methods such as Random Forests [3] are rising in popularity for genome-wide data analysis due to their ability to handle high-dimensional data, and to consider multiple predictor variables simultaneously. Random Forests (RF) trains an ensemble of multiple classification or regression trees on many bootstrap samples of subjects, handling high-dimensional data because only random

<sup>&</sup>lt;sup>§</sup>Corresponding author SJW: winham.stacey@mayo.edu RRF: freimuth.robert@mayo.edu JMB: biernacka.joanna@mayo.edu.

subsets of predictors are considered at each node of the tree. Predictions are aggregated, or bagged, across all trees in the forest, and the prediction model is evaluated using measures of prediction error (misclassification rate) or area under the receiver operating characteristic curve (AUC) in the out-of-bag subjects. Additionally, RF can be used to calculate variable importance measures, which assess a variable's overall influence on prediction and can be used to rank predictors. These measures have been shown to work well in low-dimensional genetic data to capture complex effects such as interactions among predictor variables [4], and they have been proposed as filtering or screening tools in high-dimensional data [5].

However, we previously evaluated the performance of RF with increasing numbers of genetic variants, and observed that the probability of detection of the causal variants (i.e. the proportion of times the causal variants were highly ranked) decreases more rapidly for interactions than for variants with marginal effects [6]. In fact, in high-dimensional data, both the detection probability and prediction error depend primarily on the marginal effect size, suggesting little advantage over univariate methods in high-dimensional data, particularly as a filter for the detection of interacting factors. This is because as the dimension grows, interacting variants will rarely appear in a tree together (especially if those variants do not exhibit strong marginal effects), and therefore the interaction is rarely modeled. Based on this observation, we aimed to develop an improved RF-based method for capturing complex models, Weighted Random Forests (wRF). The original RF gives each tree in the forest equal weight during aggregation. However, up-weighing the class votes from the better performing trees may have the potential to improve the overall predictive performance, assuming that the better performing trees are more likely to contain the causal variables.

Previously, other weighting methods have been proposed with ensemble classification methods such as RF. For example, because the RF classifier tends to be biased towards the majority class, weights can be assigned to each class in order to deal with the issue of imbalance [7]; these class weights are applied to the node splitting criterion and the aggregation procedure in order to ensure that the minority class is weighted equally. This method is currently available in the R package 'randomForest'. In addition to class weights, random weighting of variables/attributes within each node during the construction of decision trees has been implemented in the Random Feature Weights (RFW) method [8]. The method is similar to RF; however, rather than considering a random subset of predictors at each node, RFW considers all variables with those with higher weights being more likely to be used in tree construction. Similarly, Enriched RF alters the tree-fitting procedure by using weighted random sampling to choose the subset of variables to be considered at each node; rather than being random, variable weights are assigned based on marginal q-values [9].

Rather than weighting either classes or attributes/variables, the Weight-Adjusted Voting for Ensembles (WAVE) method uses iterative weights for both samples and trees, as an extension of both bagging and boosting for ensemble methods [10]. Trees/classifiers that perform best on the hard-to-classify instances receive the strongest weight. Other treeweighting methods that have been incorporated into RF include an approach that weighs trees based on the average margin of similar instances in the OOB data [11]. The Tree-Weighting Random Forests (TWRF) proposed by Li et al implements weighted voting with weights based on tree accuracy in the OOB data [12]. Both of these methods construct and test their weights on the OOB data, which is expected to bias PE estimates. Additionally, different weighting schemes have not been compared and weighted measures of variable importance have not been developed.

In the current study, we introduce and describe the weighted Random Forest (wRF) method, which incorporates tree-level weights into the usual RF algorithm to emphasize more accurate trees in prediction and calculation of variable importance. We consider different tree-weights, and present simulations to compare the performance of wRF to the traditional RF algorithm both in terms of prediction accuracy and performance of variable importance measures. The simulations assess performance of wRF over a range of effect sizes and complex genetic models involving interactions. Lastly, we apply wRF to data from a geneset from the Study of Addiction: Genetic and Environment (SAGE)[13].

## Methods

#### Weighted Random Forests

Assume that the data consist of a binary response variable coded 0 or 1, and p predictor variables collected on N samples. The traditional Random Forests (RF) method would build an ensemble of *ntree* classification trees to predict the outcome from the predictors, with each tree trained on a different bootstrap sample of N subjects, and a random subset of *mtry* predictors considered at each node of the tree. The original implementation of RF then aggregates tree-level results equally across trees. We implement the usual RF algorithm to build the trees of the forest; however, we utilize performance-based weights for tree aggregation. In particular, we consider weighting class 'votes' from each tree in the forest such that better performing trees are weighted more heavily.

Because weights are performance-based, applying the weights to the same dataset from which the weights were calculated (as was done in [11, 12]) would bias prediction error assessment. To avoid this bias, we first split the data into training and testing sets, and use the training data to implement the usual RF algorithm, with trees constructed on *ntree* bootstrap samples. Using the out-of-bag (OOB) individuals, estimates of the predictive ability of each tree are calculated (such as tree-level prediction error), which can be used to compute weights,  $w_j$ , for each tree j=1,...,ntree. In our implementation of wRF, the training data included <sup>3</sup>/<sub>4</sub> of the original sample ( $M_I$  individuals); thus, for each tree, approximately <sup>1</sup>/<sub>2</sub> of the full sample was in-bag and was used to build the tree, and <sup>1</sup>/<sub>4</sub> was out-of-bag and was used to assess tree performance and calculate tree weights.

After the tree-weights are computed in the training data, we use the *ntree* trees to obtain votes for the <sup>1</sup>/<sub>4</sub> observations in the independent test data, and aggregate the votes (predicted classifications) across trees by applying the weights  $w_j$  to the votes. Let  $v_{test,ij}$  be the vote for tree *j* for subject *i* in the independent test data, where *i*=1,...,  $M_2$ =N/4. Then the weighted prediction for subject *i* based on all trees is:

$$wP_i = \sum_{j=1}^{ntree} w_j \cdot v_{test,ij} \quad (1)$$

Using the weighted predictions in (1), we can compute performance measures of the weighting procedure in the independent test set, such as the prediction error ( $PE_{wRF}$ ) and AUC ( $AUC_{wRF}$ ) of the Weighted Random Forest. If  $y_i$  is the true class of subject *i*, then the prediction error  $PE_{wRF}$  can be computed based on the weighted classifications  $wC_i$ :

$$wC_{i} = I (wP_{i} \ge 0.5)$$
$$PE_{wRF} = \frac{1}{M_{2}} \sum_{i=1}^{M_{2}} |wC_{i} - y_{i}| \quad ^{(2)}$$

Similarly, we can use the weighted predictions  $wP_i$  (Equation 1) to calculate the weighted AUC ( $AUC_{wRF}$ ).

Note that setting  $w_j=1/ntree$  gives the usual aggregate prediction in the traditional implementation of RF, and then  $PE_{wRF}$  is equivalent to the usual estimate of PE computed on the test set.

In addition to prediction and PE assessment, the usual RF algorithm can be used to estimate the importance of each variable used in construction of the classifier. The most commonlyused estimate of variable importance is permutation importance, or mean decrease in accuracy (MDA)—the increase in prediction error (or reduction in prediction accuracy) after the data for the variable of interest are permuted across all individuals within each tree [14]. Based on (1), we can also compute a weighted version of the MDA variable importance:

$$VI_{wRF} = \frac{1}{M_2} \sum_{i=1}^{M_2} |wP_i^* - y_i| - |wP_i - y_i| \quad (3)$$

If  $w_i = 1/ntree$ ,  $VI_{wRF}$  is equivalent to the usual MDA variable importance.

#### **Choice of Weights**

To up-weigh better performing trees, weights  $w_j$  should be chosen based on some measure of predictive ability at the tree-level. Intuitively, weights inversely related to tree-level prediction error in the OOB training data are appropriate. In the OOB training data, define  $v_{train,ij}$  as the vote for subject *i* in tree *j* and let *oob*<sub>ij</sub> be an indicator for the out-of-bag status of subject *i* in tree *j*. We define tree-level prediction error for tree *j* as:

$$tPE_{j} = \frac{1}{\sum_{i=1}^{M_{1}} oob_{ij}} \sum_{i=1}^{M_{1}} |v_{train,ij} - y_{i}| \cdot oob_{ij}$$
(5)

In our proposed wRF implementation, we utilize weights of the form  $w_j = x_j / \sum_{j=1}^{ntre} x_j$ , such that  $\sum_{j=1}^{ntree} w_j = 1$ . To implement the usual RF aggregation, let  $x_j = 1$ ; however, weights such as  $x_j = 1 - tPE_j$  or  $x_j = \frac{1}{tPE_j}$  will allow for a higher weight for more accurate trees.

Additionally, we may wish to further up-weigh the best performing trees by utilizing

weights with a right skewed distribution, such as  $x_j = \exp\left(\frac{1}{tPE_j}\right)$  or  $x_j = \left(\frac{1}{tPE_j}\right)^{\lambda}$  for some  $\lambda$ . In fact, this strategy is supported by previous work [8], which showed that skewed weights improved performance. Alternatively, it may be advantageous to increase variability across weights by considering weights with a uniform distribution, such as

$$x_j = rank\left(\frac{1}{tPE_j}\right)$$

In addition to weights based on  $tPE_j$ , other measures of tree-level performance could be used. For instance, to capture the correlation between tree-level predictions and observed responses, we consider regression-based weights constructed from the estimated regression coefficient  $\hat{\beta}_i$  from the following linear model fit to the OOB subjects from tree *j*:

$$y_i = \alpha_j + \beta_j v_{train,ij} + \varepsilon$$
 (6)

Intuitively,  $\hat{\beta}_j$  will be larger if the predicted votes for tree *j* are close to the true responses, and close to 0 or less than 0 if the predicted votes are either uncorrelated or negatively

correlated with the true responses. This suggests a weight such as  $x_j = \hat{\beta}_j + \min|\hat{\beta}_j|$ , shifted such that all  $x_j > 0$ .

#### Simulation Study

In order to compare the predictive performance of wRF to RF with equal tree-weights and to evaluate various choices of tree-weights, we conducted a simulation study. Because the goal of this work was to explore the potential improvement of wRF over RF for the analysis of high-dimensional genetic data, we simulated datasets consisting of p=1000 predictors representing genotypes at single nucleotide polymorphisms (SNPs), with the genotypes (predictors) coded as (0,1,2) representing the number of copies of the variant allele that a person carries at the SNP. Datasets were designed to reflect a case/control study with N=1000 subjects, including 500 cases and 500 controls. For simplicity, SNP genotypes were generated assuming a fixed minor allele frequency of 0.3 and independence across predictor variables, in order to avoid confounding effects of varying predictor frequencies or correlation among predictors. Conditional on the genetic variants, an intermediate quantitative response variable Q was generated to reflect an underlying quantitative trait, caused by a series of *m* causal pairs of SNPs with possible pairwise interactions. We assumed that Q is actually unobserved, and instead we observe the binary manifestation Y. Q was simulated under the following Gaussian model:

$$Q = \beta_0 + \sum_{k=1}^{m} \left( \beta_1 X_{1,k} + \beta_2 X_{2,k} + \beta_3 X_{1,k} X_{2,k} \right) + \varepsilon \quad (7)$$

where  $\varepsilon \sim N(0, \sigma)$  and  $X_{i,k}$  refers to the genotype (0,1,2) for *i*th SNP of the *k*th SNP pair. Our model for Q represents a flexible class of models, where the number of causal pairs, effect sizes, and strength or presence of interactions can vary to reflect different genetic scenarios depending on parameter choices. In our simulations, m=2 or 10 causal SNP pairs, yielding 2m causal predictor variables and p-2m variants that are not associated with the underlying quantitative trait Q. The binary response Y, representing disease status, was generated based on a threshold model, where Y=I(Q > median(Q)).

The parameter  $\beta = [\beta_1, \beta_2, \beta_3]$  controls the effect sizes for the genetic model, and was chosen to achieve a desired total broad sense heritability ( $H^2$ ), the proportion of trait variation that can be explained by genetic variation [6, 15]. Broad sense heritability ( $H^2$ ) was utilized to quantify the overall effect size of a particular genetic model and the contribution of a variable due to both marginal and interaction components. Suppose a binary disease trait is controlled by two genetic variants A and B. Then the total heritability due to the two variants ( $H^2_{AB}$ ) can be partitioned into the marginal effect of each variable ( $H^2_{M,A}, H^2_{M,B}$ ) and the interaction effect ( $H^2_{I,AB}$ ), representing the deviation from additivity:

$$H_{AB}^2 = H_{M,A}^2 + H_{M,B}^2 + H_{I,AB}^2$$

In this study,  $\beta$  was chosen such that  $H^2$  ranged from 0 to 20%, with equal effects  $H^2/m$  per pair of causal variants, and the ratio between the marginal effects  $(H^2_M)$  and the interaction effects  $(H^2_I)$  of each pair of causal variants was 0.5, 1, or 2. As a baseline comparison, we also simulated data with only marginal effects (i.e.  $H^2_{I,AB}=0$  and  $\beta_3=0$ ) for the scenarios

with m=10 causal loci. Specific values of  $\beta$  and  $H^2$  are shown in Tables 1 and 2, giving the range of simulated scenarios for the class of models investigated in this study.

For each combination of m,  $H^2$  and ratio  $H^2_M/H^2_I$  (resulting in 34 simulation scenarios), 100 datasets were generated and analyzed using wRF with *ntree*=5000 and *mtry*=sqrt(*p*), the default value for the number of predictor variables to be considered at each node. Each dataset was analyzed with the previously described weights:

 $x_i=1$  (which yields the original RF without weighting),

$$1 - tPE_{j},$$

$$\exp\left(\frac{1}{tPE_{j}}\right),$$

$$\left(\frac{1}{tPE_{j}}\right)^{\lambda}, \text{ where } \lambda=1,2,3,4,5$$

$$rank\left(\frac{1}{tPE_{j}}\right),$$

$$\hat{\beta}_{j}+\min|\hat{\beta}_{j}|, \text{ based on Equation 6.}$$

For each simulation replicate,  $M_1$ =750 of the 1000 subjects made up the training set, and  $M_2$ =250 made up the test set. Rather than using the usual OOB predictions to assess unweighted PE from the standard RF approach, weights  $x_j$ =1 were used to represent RF analysis without weighting, so that all results were obtained from the same forest and evaluated on the same test set under the wRF framework, making the weighted and unweighted results more comparable. Performance was measured in terms of  $PE_{wRF}$ ,  $AUC_{wRF}$ , and ranks based on  $VI_{wRF}$  of the causal variants. All analyses were performed in R statistical software.

#### **Real Data Application**

To assess the performance of wRF in a real dataset, we applied both the traditional RF method along with wRF to a candidate gene-set from a genome-wide association study of alcoholism, the Study of Addiction: Genetic and Environment (SAGE)[13], available on Database of Genotypes and Phenotypes (dbGaP)[16]. We investigated associations with SNPs from the NMDA-dependent AMPA trafficking cascade pathway. Previously, using traditional regression-based methods, we showed that this gene-set was significantly associated with alcohol dependence (p<0.01) [17]. Because RF requires complete data, we utilized an imputed dataset, and excluded SNPs from the X chromosome to avoid confounding with sex. Thus, after quality control, the analyzed dataset consisted of genotypes of 1149 cases and 1357 controls at 711 SNPs in 8 genes, including: *GRIN1*, *GRIN2A*, *GRIN2B*, *CAMK2A*, *CAMK2B*, *GRIA1*, *GRIA2*, and *GRIA4*.

We ran wRF with all weights discussed here, including equal weights (equivalent to traditional RF with an independent test set used for evaluation of prediction). Data were split 75% for training and 25% for testing, as in our simulations. We present results for a single data-split, as well as 4-fold cross-validation results to assess the sensitivity of the weighted analysis to a particular random split. For comparability, we assess analysis with wRF with and without the use of equal tree-weights. We also conducted a traditional RF analysis (using the usual OOB estimates) and compare the results to wRF, although it should be noted that the usual OOB prediction estimate is biased [18, 19].

#### Results

#### Simulation Study

For the simulation study with two causal pairs of SNPs, performance in terms of both  $PE_{wRF}$  and  $AUC_{wRF}$  improved as H<sup>2</sup> (i.e. overall genetic effect size) increased. Results are plotted in Figure 1A and B for the weights  $x_j=1,(1/tPE_j)^2$ ,  $(1/tPE_j)^5$ , and rank $(1/tPE_j)$ ; results for the other weights followed a similar pattern and can be seen in Supporting Information (Supporting File 1). For low H<sup>2</sup>, performance was poor with  $PE_{wRF}$  and  $AUC_{wRF}$  both near 0.5, and all weighting methods were nearly identical. As H<sup>2</sup> increased,  $PE_{wRF}$  decreased and the various types of weights displayed more variation in performance. For high heritability and for all ratios of marginal to interacting effects (H<sup>2</sup><sub>M</sub>/H<sup>2</sup><sub>I</sub>), equal tree-weights yielded the largest  $PE_{wRF}$  estimates, while the lowest estimates of  $PE_{wRF}$  were observed for the most

extreme tree-weights, such as  $x_j = rank \left(\frac{1}{tPE_j}\right)$  and  $x_j = \left(\frac{1}{tPE_j}\right)^5$ . However, the advantage was not substantial; the largest improvement was observed in the case of stronger interaction effects (H<sup>2</sup><sub>M</sub>/H<sup>2</sup><sub>I</sub>=0.5) for H<sup>2</sup>=15%, where there was approximately a 2.5%

improvement in  $PE_{wRF}$  by use of  $x_j = \left(\frac{1}{tPE_j}\right)^5$  compared to equal tree-weights (Figure 1A). When  $AUC_{wRF}$  was used to measure performance, a similar pattern was observed as H<sup>2</sup> increased (Figure 1B).

For the simulations involving 10 causal interacting SNP pairs, we observed similar results, although because the same total effect size is dispersed across more pairs of SNPs, the predictive ability was generally lower. Results are plotted in Figure 2A and B for weights  $x_j=1,(1/tPE_j)^2,(1/tPE_j)^5$ , and rank $(1/tPE_j)$ ; results for the other weights were in the same range and are shown in the Supporting Information (Supporting File 2). When H<sup>2</sup>=5%, performance with  $PE_{wRF}$  and  $AUC_{wRF}$  was nearly identical for all weighting methods. Performance became more differentiated as H<sup>2</sup> increased (Figure 2A and B), with the estimated  $PE_{wRF}$  being consistently highest for  $x_j=1$ , although the difference between results with different weights was very small. For scenarios with high total heritability (and all values of H<sup>2</sup><sub>M</sub>/H<sup>2</sup><sub>I</sub>, including scenarios with H<sup>2</sup><sub>I</sub>=0), the lowest  $PE_{wRF}$  estimates were

observed for the uniformly-distributed tree-weights,  $x_j = rank\left(\frac{1}{tPE_j}\right)$ , although the largest gain over the currently implemented equal weighting was only about 0.5% (Figure 2A). Results in terms of  $AUC_{wRF}$  were consistent with those of  $PE_{wRF}$ ; an improvement observed via weighting was very small (Figure 2B). Furthermore, the ratio of marginal to interacting effect sizes ( $H^2_M/H^2_I$ ) seemed to have little impact on performance; the pattern of results was similar for scenarios with only marginal effects ( $H^2_I=0$ ) as for scenarios with stronger interactions ( $H^2_M/H^2_I=0.5$ ); see Supporting Information, Supporting File 2. Rather, the total genetic effect size ( $H^2$ ), as well as the per SNP effect size, seemed to be the driving factor on performance of wRF.

For both sets of simulation studies with either m=2 or 10 causal pairs of SNPs, there was very little change amongst weighted variable importance measures for the causal factors (Supporting Information, Supporting Files 1 and 2). Variable importance measures are typically used to rank variables, therefore we expected that the causal SNPs would rank highly out of all p=1000 predictor variables. To compare  $VI_{wRF}$  measures across all causal SNPs, we computed the average ranks across all 2\*m causal variants. For each scenario, average ranks were almost indistinguishable across choice of weights, with the exception of the uniform-weights which produced slightly lower  $VI_{wRF}$  average ranks, as can be seen in Supporting Information. Median ranks demonstrated similar results.

#### **Real Data Application**

Weighted RF was fit to the 711 SNPs in the NMDA-dependent AMPA trafficking cascade pathway, with the same set of weights that were investigated in the simulation study, including  $x_j$ =1 representing an unweighted RF. Overall, the SNPs were not highly predictive of alcohol dependence; PE ranged from 0.476-0.483 and AUC ranged from 0.503-0.521. Results of the traditional analysis using RF with the OOB estimates yielded PE and AUC very similar to those seen with all of the weights. Variable importance measures differed slightly between the different methods, with rs980272 near CAMK2A and rs2267779 and rs2267780 in GRIN2A consistently ranking near the top of the results in the wRF analyses; rs1421109 and rs12322168 in GRIN2B ranked at the top of the traditional RF analyses. Results for wRF were similar between the single split of the data and cross-validation for PE/AUC, although some differences are observed for  $VI_{wRF}$ ; however, estimates of variable importance are known to be unstable[20]. Results are shown for weights  $x_j$ =1,(1/tPE\_j)<sup>2</sup>, (1/tPE\_i)<sup>5</sup>, and rank(1/tPE\_i) (Table 3; Figure 3; Supporting Information, Supporting File 3).

## Discussion

In this study, we proposed Weighted Random Forests, an extension of Random Forests motivated by the poor performance of RF to detect interactions in high-dimensional genetic data. The wRF method weighs better performing trees more heavily during aggregation. Our simulation studies demonstrated that the performance of wRF (as measured by  $PE_{wRF}$  and  $AUC_{wRF}$ ) is at least as good as RF with equal tree-weights, and in some situations the predictive capability is slightly improved. For instance, when the effect size of the causal variants is large (i.e. high H<sup>2</sup> per causal SNP pair) then wRF has lower prediction error. The improvement in prediction is greater for the strongest weights with the greatest variability

(i.e.  $x_j = rank\left(\frac{1}{tPE_j}\right)$  and  $x_j = \left(\frac{1}{tPE_j}\right)^5$ ). This is in agreement with previous results showing that skewed weights of the form  $w^{\lambda}$  improved performance [8].

However, the gain in predictive ability is quite modest, particularly for data with strong interactions among predictor effects. We only observe meaningful improvements in prediction accuracy when effect sizes are large (i.e.  $H^2 > 15\%$ )—sizes that are rarely observed in genetic epidemiologic studies. In scenarios with weak effects, the results of wRF are essentially the same as RF; aggregation with equal weighting across trees produces similar results as aggregation with weights with a high degree of variability (such as highly skewed weights or weights with a uniform distribution). Although performance of wRF is slightly better in situations where the interaction effects are stronger than the marginal effects ( $H^2_M/H^2_I < 1$ ), the gain in predictive accuracy is small in these situations. Overall, the wRF method performs similarly to the traditional RF method, and there is little advantage given the added complexity of introducing performance-based weights.

We applied the wRF method to a real genetic dataset similar in size to those evaluated in the simulation studies. The performance of wRF on this dataset is consistent with the results of the simulation studies. Generally, the analyzed SNPs were not highly predictive of alcohol dependence, generating high PE estimates. In this situation, analysis with wRF with and without equal weights produces similar predictive performance; likewise, the traditional analysis using RF with the usual OOB did not yield better predictions. On the other hand, variable importance measures differ across methods. The wRF method identified rs980272 in CAMK2A as being one of the most important SNPs, which is consistent with previously published results [17]. Top ranking SNPs based on the traditional RF analysis were not consistent with the previously reported single SNP results.

Given our intuitive motivation for the method, it may seem surprising that we do not observe a greater improvement with wRF. In genetic association studies, the expected effect sizes are modest, as complex genetic traits are influenced by a large number of causal variants with very small effect (i.e.  $H^2 < 1\%$  per variant). Because the effect sizes are very small, even the most predictive trees within the forest are still not very accurate, and variation in predictive performance from tree to tree is low. However, the performance of wRF may be improved for other applications outside of genetics, where larger effect sizes that induce greater tree variability can be expected.

The advantages of wRF are possibly limited by the need to split the sample to avoid biased estimates of PE and variable importance. In particular, splitting the full sample into independent training and testing sets may be disadvantageous due to the reduction in sample size used for tree-building. This is of particular concern for small datasets, which will be sensitive to the particular split into training and testing sets. To circumvent this limitation, other internal validation techniques could be considered. One possibility is k-fold cross-validation, where wRF is fit to k subsets of the data. We implemented this strategy in the analysis of the addiction dataset; our real dataset, like our simulated datasets, was large with 711 variables on 2506 subjects. The use of cross-validation yielded similar results to a single split in this case, although this is not likely to be true for a small dataset.

To evaluate the effect on RF and wRF performance due to the reduced sample size used for training, for a subset of our simulation scenarios we compared the results of wRF trained on 750 samples to RF trained on 1000 samples, with prediction evaluated on the same independent set of 250 individuals. The use of 1000 (as opposed to 750) samples for training improved PE and AUC estimates by about 1%. In traditional RF analysis OOB error estimates are usually used for assessment of PE, rather than an independent test set. However, OOB estimates of PE are biased in classification problems [18, 19], which complicates a direct comparison of wRF to the traditional RF implementation. Our simulations showed that this bias is conservative (~2-3% worsening of PE) and is largest for datasets with higher predictive accuracy, which is also when we see the largest improvement with the use of weighting. Although an extremely important issue, further examination of this bias is beyond the scope of this study.

Our results are consistent with those observed in the comparison of RF to the WAVE algorithm [10], which applies weights to both tree classifiers and hard-to-classify individuals, and was shown to have similar performance to Random Forest. On the other hand, the weighted voting method that computes tree-weights based on margin of similar instances [11] and the TWRF method [12] showed improved performance over traditional RF with the use of weighting. However, both of these methods were tested on low dimensional real datasets with high RF prediction accuracy, rather than high-dimensional data. Furthermore, both methods computed and tested the weights on OOB data; therefore the observed improvement was likely (at least partly) due to bias from over-fitting. We eliminated this bias from our assessment of wRF by reserving an independent test set for predictive evaluation.

The current study was designed to compare wRF with the traditional RF implementation under scenarios that reflect genetic association data with multiple, and possibly interacting, causal variants. Although beyond the scope of this study, a direct comparison with other weighting methods (such as WAVE, TWRF, etc.) would be useful. While wRF did not yield improved predictions for the weights examined here, other weighting schemes may improve performance of wRF. An important consideration in any RF analysis is the optimization of the parameters *mtry* and *ntree*. Because the focus of this study was on a comparison of the same RF algorithm under different weighting schemes, the values of *mtry*=sqrt(*p*) and

*ntree*=5000 were used for each run to aid in comparability. Adaptively tuning these parameters to each dataset, may lead to improved predictive ability of wRF.

In summary, the wRF method, which incorporates tree-level weights, does not dramatically improve predictive ability in high-dimensional genetic data, but it may improve performance in other domains that exhibit stronger effects.

## **Supplementary Material**

Refer to Web version on PubMed Central for supplementary material.

#### Acknowledgments

This study was funded by NIDA (R21 DA019570, PI Biernacka).

The datasets used for the analyses described in this manuscript were obtained from the database of Genotypes and Phenotypes (dbGaP; study accession phs000092.v1.p1). We also acknowledge the Study of Addiction: Genetics and Environment (SAGE), which is part of the Gene Environment Association Studies (GENEVA) initiative supported by the National Human Genome Research Institute

#### References

- McCarthy MI, Abecasis GR, Cardon LR, Goldstein DB, Little J, Ioannidis JP, Hirschhorn JN. Genome-wide association studies for complex traits: consensus, uncertainty and challenges. Nature reviews Genetics. 2008; 9(5):356–369.
- Manolio TA, Collins FS, Cox NJ, Goldstein DB, Hindorff LA, Hunter DJ, McCarthy MI, Ramos EM, Cardon LR, Chakravarti A, et al. Finding the missing heritability of complex diseases. Nature. 2009; 461(7265):747–753. [PubMed: 19812666]
- 3. Breiman L. Random Forests. Mach Learn. 2001; 45:5-32.
- 4. Lunetta KL, Hayward LB, Segal J, Van Eerdewegh P. Screening large-scale association study data: exploiting interactions using random forests. BMC genetics. 2004; 5(1):32. [PubMed: 15588316]
- Goldstein BA, Hubbard AE, Cutler A, Barcellos LF. An application of Random Forests to a genome-wide association dataset: methodological considerations & new findings. BMC genetics. 2010; 11:49. [PubMed: 20546594]
- Winham SJ, Colby CL, Freimuth RR, Wang X, de Andrade M, Huebner M, Biernacka JM. SNP interaction detection with random forests in high-dimensional genetic data. BMC Bioinformatics. 2012; 13(1):164. [PubMed: 22793366]
- 7. Chen C, Liaw A, Breiman L. Using Random Forest to Learn Imbalanced Data. Technical Report 666 Statistics Department of University of California at Berkley. 2004 In:
- 8. Maudes J, Rodriguez JJ, Garcia-Osorio C, Garcia-Pedrajas N. Random feature weights for decision tree ensemble construction. Information Fusion. 2012; 13(1):20–30.
- Amaratunga D, Cabrera J, Lee YS. Enriched random forests. Bioinformatics. 2008; 24(18):2010– 2014. [PubMed: 18650208]
- 10. Kim H, Moon H, Ahn H. A weight-adjusted voting algorithm for ensembles of classifiers. Journal of the Korean Statistical Society. 2011; 40(4):437–449.
- Robnik-Sikonja M. Boulicaut JF, Esposito F, Giannoti F, Pedreschi D. Improving random forests. Machine Learning: Ecml 2004, Proceedings. 2004; 3201:359–370. In:
- Hong Bo, L.; Wei, W.; Hong Wei, D.; Jin, D. Trees Weighting Random Forest Method for Classifying High-Dimensional Noisy Data; e-Business Engineering (ICEBE), 2010 IEEE 7th International Conference on: 10-12 Nov. 2010; 2010. p. 160-163.In:
- Bierut LJ, Agrawal A, Bucholz KK, Doheny KF, Laurie C, Pugh E, Fisher S, Fox L, Howells W, Bertelsen S, et al. A genome-wide association study of alcohol dependence. Proceedings of the National Academy of Sciences of the United States of America. 107(11):5082–5087. [PubMed: 20202923]

- Bureau A, Dupuis J, Falls K, Lunetta KL, Hayward B, Keith TP, Van Eerdewegh P. Identifying SNPs predictive of phenotype using random forests. Genet Epidemiol. 2005; 28(2):171–182. [PubMed: 15593090]
- 15. Culverhouse R, Suarez BK, Lin J, Reich T. A perspective on epistasis: limits of models displaying no main effect. Am J Hum Genet. 2002; 70(2):461–471. [PubMed: 11791213]
- Mailman MD, Feolo M, Jin Y, Kimura M, Tryka K, Bagoutdinov R, Hao L, Kiang A, Paschall J, Phan L, et al. The NCBI dbGaP database of genotypes and phenotypes. Nat Genet. 2007; 39(10): 1181–1186. [PubMed: 17898773]
- Karpyak VM, Geske JR, Colby CL, Mrazek DA, Biernacka JM. Genetic variability in the NMDAdependent AMPA trafficking cascade is associated with alcohol dependence. Addict Biol. 2012; 17(4):798–806. [PubMed: 21762291]
- Biau G, Devroye L, Lugosi G. Consistency of Random Forests and Other Averaging Classifiers. Journal of Machine Learning Research. 2008; 9:2015–2033.
- Bylander T. Estimating generalization error on two-class datasets using out-of-bag estimates. Mach Learn. 2002; 48(1-3):287–297.
- Boulesteix AL, Slawski M. Stability and aggregation of ranked gene lists. Brief Bioinform. 2009; 10(5):556–568. [PubMed: 19679825]



#### Figure 1.

Simulation Results (wPE and wAUC), m=2 causal pairs. Weighted Prediction Error (top row) and AUC (bottom row) plotted against total heritability for models with 2 causal interacting pairs. Results are only displayed for a subset of weights and for  $H^2_M/H^2_I = 0.5$ , 2.0; for the full results, see Supporting Information. The results for weights that are not plotted are similar and in the same range.



#### Figure 2.

Simulation Results (wPE and wAUC), m=10 causal pairs. Weighted Prediction Error (top row) and AUC (bottom row) plotted against total heritability for models with 10 causal interacting pairs. Results are only displayed for a subset of weights and for  $H^2_M/H^2_I = 0.5$ , 2.0; for the full results, see Supporting Information. The results for weights that are not plotted are similar and in the same range.



#### Figure 3.

Variable importance results from real data analysis of the NMDA-dependent AMPA trafficking cascade pathway.

Importance is plotted for each SNP by chromosomal location for A: traditional RF using OOB estimates, and wRF with cross-validation and weights of B: x=1, C:  $x=(1/tPE)^2$ , D:  $x=(1/tPE)^5$ , and E: x=rank(1/tPE). Colors indicate gene membership, as specified on the x-axis. Only SNPs with importance > 0 are plotted.

Importance is plotted for each SNP by chromosomal location for A: traditional RF using OOB estimates, and wRF with a single data split and weights of B: x=1, C:  $x=(1/tPE)^2$ , D:  $x=(1/tPE)^5$ , and E: x=rank(1/tPE). Colors indicate gene membership, as specified on the x-axis. Only SNPs with importance > 0 are plotted.

#### Table 1

Simulated effect sizes for m=2 causal pairs.

<b>H</b> <sup>2</sup>	$\mathrm{H}^{2}_{\mathrm{pair}}$	${\rm H^2}_{\rm M}/{\rm H^2}_{\rm I}$	${\rm H^2}_{\rm M}$	${\rm H^2}_{\rm I}$	β1	β2	β3
		2	0.167	0.083	0.495	0.495	0.395
		1	0.125	0.125	0.520	0.520	0.490
0.5	0.25	0.5	0.083	0.167	0.505	0.505	0.545
		2	0.333	0.167	0.710	0.710	0.570
		1	0.250	0.250	0.720	0.720	0.680
1.0	0.50	0.5	0.167	0.333	0.720	0.720	0.780
		2	0.833	0.417	1.102	1.102	0.885
		1	0.625	0.625	1.113	1.113	1.071
2.5	1.25	0.5	0.417	0.833	1.151	1.151	1.250
		2	1.667	0.833	1.583	1.583	1.276
		1	1.250	1.250	1.652	1.652	1.562
5.0	2.50	0.5	0.833	1.667	1.662	1.662	1.802
		2	3.333	1.667	2.342	2.342	1.899
		1	2.500	2.500	2.447	2.447	2.317
10.0	5.00	0.5	1.667	3.333	2.467	2.467	2.682
		2	5.000	2.500	3.022	3.022	2.468
		1	3.750	3.750	3.180	3.180	3.016
15.0	7.50	0.5	2.500	5.000	3.198	3.198	3.484

#### Table 2

Simulated effect sizes for m=10 causal pairs.

H <sup>2</sup>	${ m H}^2_{\ pair}$	${{{H}^{2}}_{M}}  /  {{H}^{2}}_{I}$	${\rm H}^2{}_{\rm M}$	${\rm H^2}_{\rm I}$	β1	β2	β <sub>3</sub>
		Inf	0.5	0	0.307	0.307	0
		2	0.333	0.167	0.482	0.482	0.386
		1	0.250	0.250	0.502	0.502	0.474
5.0	0.5	0.5	0.167	0.333	0.506	0.506	0.548
		Inf	1.0	0	0.436	0.436	0
		2	0.667	0.333	0.686	0.686	0.550
		1	0.500	0.500	0.714	0.714	0.673
10.0	1.0	0.5	0.333	0.667	0.718	0.718	0.777
		Inf	1.5	0	0.536	0.536	0
		2	1.000	0.500	0.842	0.842	0.675
		1	0.750	0.750	0.876	0.876	0.827
15.0	1.5	0.5	0.500	1.000	0.883	0.883	0.956
		Inf	2.0	0	0.622	0.622	0
		2	1.333	0.667	0.978	0.978	0.785
		1	1.000	1.000	1.016	1.016	0.960
20.0	2.0	0.5	0.667	1.333	1.024	1.024	1.109

#### Table 3

Real data analysis of the NMDA-dependent AMPA trafficking cascade pathway, PE and AUC

	RF OOB	x=1	x=(1/tPE) <sup>2</sup>	x=(1/tPE) <sup>5</sup>	x=rank(1/tPE)
PE, 1 split	0.472	0.467	0.470	0.475	0.483
PE, CV	0.479	0.467	0.468	0.467	0.465
AUC, 1 split	0.505	0.507	0.506	0.505	0.503
AUC, CV	0.512	0.521	0.521	0.522	0.520